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C.L.E.D. Medium

DM110

Intended Use
A non-inhibitory differential medium for the investigation of urinary infections.

Contents
See pack label.

Formulation*

Material:	Concentration in medium:
Peptone mixture	13.85g/litre
Lactose	8.5g/litre
L-Cystine	0.128g/litre
L-Cysteine	0.4g/litre
Bromothymol blue	0.02g/litre
Agar	15.0g/litre
Final pH: 7.3 ± 0.2	

Storage and shelf life
All dehydrated culture media containers should be kept tightly closed and stored in a dry place at 10 to 25°C until the expiry date shown on the pack label.

Precautions
For *in vitro* diagnostic use only. Observe approved hazard precautions and aseptic techniques. To be used only by adequately trained and qualified laboratory personnel. Sterilise all biohazard waste before disposal. Refer to Product Safety Data sheet (available on request or via MAST® website).

Materials required but not provided
Standard microbiological supplies and equipment such as loops, MAST® selective supplements, swabs, applicator sticks, incinerators and incubators, etc., as well as serological and biochemical reagents and additives such as blood.

Procedure
1. Suspend by swirling 37.9g of powder in 1 litre of distilled or deionised water.
2. Autoclave at 121°C (15 p.s.i.) for 15 minutes.
3. Mix thoroughly, pour culture plates (15 to 20ml per plate) and allow to set.
4. Prepared culture plates may be used immediately or stored in plastic bags at 2 to 8°C for up to one week before use.
5. Inoculate plates with early-morning urine (EMU), mid-stream urine (MSU) or catheter urine (CSU) by surface plating, streaking out for single colonies. Alternatively, colony counts can be obtained by spreading evenly over the entire surface. MAST® C.L.E.D. medium can be used in conjunction with MAST® BACTERURITEST Strips (BTR1) for the screening of urine cultures.
6. Incubate plates aerobically for 18 to 24 hours at 35 to 37°C

Interpretation of results
After incubation, record growth of organisms. Typical characteristics to note include colony size, morphology, pigmentation and effect on surrounding medium. Lactose is included as a carbon source, therefore lactose and non-lactose fermentors can be easily differentiated by a colour change of the medium - organisms capable of fermenting lactose will lower the pH and turn the medium yellow.

Quality control
Check for signs of deterioration. Quality control must be performed with at least one organism to demonstrate expected performance. Do not use the product if the result with the control organism is incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain.

Test Organisms	Result
Escherichia coli ATCC® 25922	Growth (Yellow)
Proteus mirabilis ATCC® 29906	Growth (Blue / green)
Staphylococcus aureus ATCC® 25923	Growth (Yellow)

References
Bibliography available on request.

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*Formulation may change to meet performance criteria