

## Actinomycete MAST® SELECTATAB

### MS25 Series

#### Intended Use

For the selective isolation of actinomycetes.

FOR IN VITRO DIAGNOSTIC USE ONLY

#### Contents

25 (small) or 10 (large) MAST® SELECTATAB.

See pack label.

#### Formulation

Material:	Concentration in medium:
Metronidazole	2.5 mg/L
Nalidixic Acid	25 mg/L

#### Storage and shelf life

Store unopened at 2 to 8°C until the expiry date shown on the pack label. Once opened, store MAST® SELECTATAB in capped, original packaging at 2 to 8°C until the expiry date shown on the pack label.

#### Precautions

For *in vitro* diagnostic use only. Observe approved biohazard precautions and aseptic techniques. To be used only by adequately trained and qualified laboratory personnel. Sterilise all biohazard waste before disposal. Refer to Product Safety Data sheet.

#### Materials required but not provided

Standard microbiological supplies and equipment such as loops, MAST® culture media, swabs, applicator sticks, incinerators and incubators, etc., as well as serological and biochemical reagents and additives such as blood.

#### Procedure

1. Label Petri dishes using the self-adhesive labels provided.
2. Sterilise the appropriate volume of MAST® Columbia Agar (DM115D), cool to 50 to 55°C and stand in a water bath at this temperature.
3. Using sterile forceps, add one MAST® SELECTATAB to the volume of medium as stated on the pack label and label the bottle. Allow to stand for several minutes at 50 to 55°C in the water bath until the MAST® SELECTATAB has broken up.
4. After the MAST® SELECTATAB has broken up, swirl 3 to 4 times and invert to complete dispersal. An alternative method is to first dissolve the MAST® SELECTATAB in 3 to 5 mL of sterile water and add this to the appropriate volume of medium.

5. Supplement the medium with 10% sterile defibrinated horse blood, mix thoroughly and pour culture plates (15 to 20ml per plate) and allow to set.
6. Prepared culture plates may be used immediately or stored in plastic bags at 2 to 8°C for up to one week before use.
7. Spread inoculate over the surface of separate dried plates 0.1mL of each sample dilution as prepared below.
8. Prepare samples for inoculation in the following manner. Place swab, coil (Intrauterine Contraceptive Device) or any pus from the coil into 5 mL MAST® Thioglycollate Broth (DM220D) and from this primary dilution make tenfold dilutions ( $10^{-1}$  to  $10^{-4}$ ) in Thioglycollate Broth. Prior dilution of the sample is important making it easier to identify small actinomycete colonies.
9. Plates should be incubated anaerobically in 10% CO<sub>2</sub> and examined after 4, 10 and 14 days.

#### Interpretation of results

*Actinomyces* spp. appear as irregular shaped white colonies approximately 2 to 4 mm in diameter, which are very adherent and difficult to emulsify.

#### Quality control

Check for signs of deterioration. Quality control must be performed with at least one organism to demonstrate a positive reaction and at least one organism to demonstrate a negative reaction. Do not use the product if the reactions with the control organisms are incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain.

Test Organisms	Result
<i>Staphylococcus aureus</i> ATCC® 25923	No growth
<i>Proteus mirabilis</i> ATCC® 29906	No growth
<i>Candida albicans</i> ATCC® 90028	Growth
<i>Bacteroides fragilis</i> ATCC® 25285	No growth
<i>Clostridium perfringens</i> ATCC® 13124	No growth
<i>Actinomyces israelii</i> ATCC® 10049	Growth

#### References

Bibliography available on request.