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## Campylobacter Enrichment (Exeter) MAST® SELECTAVIAL

### SV59 Series

#### Intended use

For the isolation of *Campylobacter* spp. from food and water samples in conjunction with Campylobacter Growth Supplement (FBP) (SV61 series)

FOR IN VITRO DIAGNOSTIC USE ONLY

#### Contents

10 vials of MAST® SELECTAVIAL.

#### Formulation

Material:	Concentration in medium:
Trimethoprim	10 mg/L
Rifampicin	5 mg/L
Polymyxin B	2500 iu/L
Cefoperazone	15 mg/L
Amphotericin B	2 mg/L

#### Storage and shelf life

Store unopened at 2 to 8°C until expiry date shown on pack label. Once reconstituted use immediately.

#### Precautions

For *in vitro* diagnostic use only. Observe approved biohazard precautions and aseptic techniques. To be used only by adequately trained and qualified laboratory personnel. Sterilise all biohazard waste before disposal. Refer to Product Safety Data sheet.

#### Materials required but not provided

Standard microbiological supplies and equipment such as loops, MAST® culture media, swabs, applicator sticks, incinerators and incubators, etc., as well as serological and biochemical reagents and additives such as blood.

#### Procedure

##### For Exeter Camp Selective Enrichment Broth

1. Sterilise the appropriate volume of Mast® Nutrient Broth (DM180D), cool to 50 to 55°C and hold at this temperature. For the preparation of agar add 15 g/L of agar to the broth before sterilisation and supplement as above.
2. Reconstitute the contents of one vial and a corresponding vial from the SV61 series, using the diluent specified on the pack label. The best method is to aseptically add the diluent using a sterile needle and syringe. Draw the diluent into the syringe and after removing the plastic cap, inject through the rubber stopper of the vial. The lyophilised supplement will rapidly dissolve and may be withdrawn into the syringe.
3. Add the antibiotic supplements to the volume of medium specified on the pack label and discard the needle into an approved container.

4. Aseptically add 5% v/v lysed horse blood and mix.
5. Prepare a 10<sup>-1</sup> homogenate of food sample using either a stomacher or blender by homogenising 25g or 25ml of sample in 225 mL of prepared broth. Incubate at 37°C for 48 hours in a tightly closed container with minimal head space.
6. For certain samples e.g. milk or water containing cold-injured *Campylobacter*, incubation at 37°C for 2 hours in a non selective broth, using FBP Growth Supplement alone, has been shown to improve isolation. After the initial 2 hour period the antibiotic supplement can be added and incubation continued.
7. Subculture onto a suitable Campylobacter selective agar such as Exeter or Preston (MAST® DM251D/MS18) medium after 24 and 48 hours. Incubate the plates in a microaerobic atmosphere at 42°C for 24 to 48 hours.

#### Interpretation of results

Examine the plates for suspect *Campylobacter* colonies - typically colonies of *C. jejuni* tend to appear as grey moist flat spreading colonies. Some strains may produce a green hue or metallic sheen. Colonies of *C. coli* tend to be creamy grey in colour, moist, slightly raised and often discrete with less spreading than *C. jejuni*.

#### Quality control

Check for signs of deterioration. Quality control must be performed with at least one organism to demonstrate a positive reaction and at least one organism to demonstrate a negative reaction. Do not use the product if the reactions with the control organisms are incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain.

Test Organisms	Result
<i>Campylobacter jejuni</i> ATCC® 33291	Growth
<i>Escherichia coli</i> ATCC® 25922	No growth

#### References

Bibliography available on request.